

A synaptic DEG/ENaC ion channel mediates learning in *C. elegans* by facilitating dopamine signalling

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An important component of learned behaviour is the ability to forecast positive or negative outcomes based on specific sensory cues. Predictive capacity is typically manifested by appropriate behavioural patterning. However, the molecular mechanisms underlying behavioural plasticity are poorly understood. *Caenorhabditis elegans* displays experience-dependent behavioural responses by associating distinct environmental signals. We find that ASIC-1, a member of the degenerin/epithelial sodium channel family, which localizes at presynaptic terminals of dopaminergic neurons, is required for associative learning in *C. elegans*. ASIC-1 functions in these neurons to amplify normal dopaminergic signalling, necessary for associative learning. Our results reveal a novel role of DEG/ENaC ion channels in neuronal communication by enhancing the activity of dopaminergic synapses. Similar mechanisms may facilitate synaptic plasticity in vertebrates.

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Introduction

The nervous system processes and integrates sensory input to generate both immediate responses and long-term changes in behaviour. Associative learning, in the form of classical and operant conditioning, allows animals to capture the logical structure of their habitat and is critical for negotiating complex environments (Pavlov, 1928; Skinner, 1930). Similar to vertebrates, the soil-dwelling nematode *Caenorhabditis elegans* integrates sensory input to predict vital field conditions, such as food availability or the presence of predators (Mohri *et al.*, 2005; Zhang *et al.*, 2005). Several paradigms of experience-dependent behavioural plasticity have been described in *C. elegans* (Nuttley *et al.*, 2002; Hobert, 2003; Rankin, 2004; Bargmann, 2005). Worms display associative learning in the form of conditioning to chemical, thermal and

mechanical cues (Gomez *et al.*, 2001; Saeki *et al.*, 2001; Law *et al.*, 2004; Mori *et al.*, 2008).

Modulation of synaptic transmission by specific ion channels localized at both pre- and postsynaptic termini facilitates synaptic plasticity, associated with learned patterns of behaviour (Hebb, 1949; Abbott and Nelson, 2000; Voglis and Tavernarakis, 2006). Acid-sensing ion channels (ASICs) have been implicated neurotransmission in the central nervous system (Wemmie *et al.*, 2006). ASICs are gated by protons and belong to the family of degenerin/epithelial sodium channels (DEG/ENaC), a large group of ion channels that are sensitive to the diuretic amiloride (Kellenberger and Schild, 2002). ASICs have been identified in diverse organisms ranging from nematodes to mammals, including humans and share a high degree of sequence and overall structure similarity (Krishtal, 2003). ASIC-1, a member of the mammalian ASIC subfamily has been identified in high synaptic density regions of the hippocampus, where it localizes at both pre- and postsynaptic membranes of synaptosomes. ASIC-1 deficiency impairs long-term potentiation (LTP) in the hippocampus, indicating that ASIC activity is critical for synaptic plasticity (Wemmie *et al.*, 2002). In addition to the hippocampus, ASIC expression has also been detected in the amygdala, one of the key brain structures involved in fear conditioning (Wemmie *et al.*, 2003). Disruption of ASIC-1 attenuates learned fear behaviour, whereas overexpression of ASIC-1 in the brain enhances fear conditioning (Wemmie *et al.*, 2004).

These findings indicate that in mammals ASICs modulate synaptic plasticity, probably by facilitating proton-evoked sodium currents at postsynaptic termini (Wemmie *et al.*, 2006). However, the underlying mechanisms linking ASIC function to learning and memory remain poorly understood. In addition, the function of ASICs at presynaptic terminals is unclear. Here, we present evidence that ASIC-1, a new member of the DEG/ENaC family, contributes to sustain enhanced dopaminergic neurotransmission, necessary for associative learning in *C. elegans*. We found that ASIC-1 localizes at presynaptic terminals of dopaminergic neurons. Animals lacking ASIC-1 are defective in associative learning, although they show normal sensory perception. Synaptic release at dopaminergic neurons is compromised in the absence of ASIC-1 activity, suggesting that modulation of dopaminergic signalling by ASIC-1 mediates associative learning in *C. elegans*. Given that ASIC ion channels are highly conserved across phyla (Krishtal, 2003), similar mechanisms may facilitate synaptic plasticity in vertebrates.

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Results

ASIC-1 is required for associative learning in *C. elegans*

To assay associative learning capacity in the nematode, we utilize two robust assays, chemosensory conditioning and

feeding-dependent thermotaxis, each based on different sets of sensory inputs. In chemosensory conditioning, animals that are normally attracted to certain water-soluble or volatile compounds avoid these chemoattractants after being subjected to starvation in their presence (Bargmann and Horvitz, 1991; Colbert and Bargmann, 1995; Saeki *et al.*, 2001). In feeding-dependent thermotaxis, animals challenged with a radial thermal gradient track the isothermal curve corresponding to their prior cultivation temperature, under *ad libitum* feeding conditions (Hedgecock and Russell, 1975; Mori, 1999; Gomez *et al.*, 2001; Mori *et al.*, 2008). We found that punishment-induced aversion towards starvation-paired chemosensory stimuli is significantly impaired in animals carrying a genetic lesion (deletion allele *ok415*; *C. elegans* Gene Knockout Consortium) in the *asic-1* gene, which encodes a member of the acid-sensing family of ion channels (Supplementary Figures 1–3). Mutant animals perform normal chemotaxis towards the compounds used, indicating that learning defects are not due to sensory perception deficiency (Figure 1A–D). We have also examined the capacity of *asic-1(ok415)* mutants for chemosensory adaptation (Bernhard and van der Kooy, 2000). We found that these animals display normal adaptation behaviour (Supplementary Figure 8). Furthermore, the feeding-dependent thermotaxis behaviour of *asic-1(ok415)* mutants is compromised (Figure 1E). Finally, although wild-type animals show enhanced chemotaxis towards soluble and volatile compounds after conditioning in the presence of food (Saeki *et al.*, 2001), *asic-1(ok415)* mutants remain unaffected (Figure 2A and B). RNAi-mediated knock down of ASIC-1 recapitulated the phenotypes of the mutant, albeit with low penetrance because of the low efficiency of RNAi in the *C. elegans* nervous system (Tavernarakis *et al.*, 2000). Other behaviours and anatomical parameters that could interfere with learning assays (locomotion, speed, mechanosensory and thermosensory responses, body size and others) are unaffected in these animals (Table I; Supplementary Figure 5 and data not shown). Thus, ASIC-1 deficiency specifically disrupts at least two different forms of associative learning in *C. elegans*.

ASCs belong to the family of DEG/ENaC, a large group of ion channels that are sensitive to the diuretic amiloride (Kellenberger and Schild, 2002) (Supplementary Figure 1). ASICS have been identified in diverse organisms ranging from nematodes to mammals, including humans and share a high degree of sequence and overall structure similarity (Krishtal, 2003) (Supplementary Figures 2 and 3). The strong conservation across species suggests that ASICS shared a common ancestor relatively early in evolution. Members of the mammalian ASIC subfamily are gated by protons. Expression of these channels in heterologous cells generates transient, H⁺ gated, amiloride-sensitive, sodium-selective currents (Waldmann *et al.*, 1997). *C. elegans* ASIC-1 shares extensive sequence similarity with vertebrate ASICS (Supplementary Figure 3). The deletion in the *ok415* allele removes a highly conserved extracellular, cysteine-rich domain of the protein that is essential for function (Supplementary Figures 3 and 4). Interestingly, part of the deleted region corresponds to the proton-sensing domain of vertebrate ASICS (Chen *et al.*, 2007; Jasti *et al.*, 2007; Paukert *et al.*, 2008). Introduction of the wild-type *asic-1* gene in *asic-1(ok415)* mutants restores learning capacity in these animals (Figure 2C and D).

ASIC-1 functions in dopaminergic neurons to mediate associative learning

asic-1 is expressed in eight neurons comprising the *C. elegans* dopaminergic system (Figure 3A; the four cephalic neurons; two anterior deirid neurons and two posterior deirid neurons; CEP, ADE, PDE, respectively) (Chase *et al.*, 2004; Kindt *et al.*, 2007). Expression is also detected in four additional neurons in the tail, two of which are the bilaterally symmetric PVQ interneurons (Figure 3A). We examined the subcellular localization of ASIC-1 in these neurons by generating two GFP reporter fusions. The first is a full-length ASIC-1::GFP reporter encompassing the complete ASIC-1 protein. To generate the second reporter, we fused GFP with the intracellular amino-terminal region of ASIC-1, which contains amino-acid sequence motifs sufficient for proper localization of these ion channels in *C. elegans* (ASIC-1N::GFP) (Tavernarakis *et al.*, 2001). Both the full-length ASIC-1::GFP and amino-terminal ASIC-1N::GFP chimaeras showed prominent punctate distribution along the processes of dopaminergic neurons, in synapse-rich areas (Figure 3B). A DsRED-synaptobrevin fusion colocalizes with both ASIC-1::GFP and ASIC-1N::GFP in these puncta (Figure 3C and D). Thus, ASIC-1 localizes at presynaptic termini of dopaminergic neurons.

To determine the requirement for ASIC-1 function in specific subsets of neurons, we examined populations of mosaic transgenic animals for conditioning to isoamyl alcohol. We found that expression of *asic-1* in all eight dopaminergic neurons is both necessary and sufficient to restore normal learning capacity in *asic-1(ok415)* mutants (Table II). Partial expression in the two ADEs or the four CEPs or the two PDEs (the three different classes of nematode dopaminergic neurons) results in intermediate rescue. We conclude that ASIC-1 function in all dopaminergic neurons contributes to normal conditioning behaviour.

Dopaminergic signalling is required for conditioning

ASCs are widely distributed in the mammalian brain, where they have been implicated in the phenomena of synaptic plasticity at postsynaptic ends, and learning and memory (Wemmie *et al.*, 2002, 2006). What function does ASIC-1 serve at presynaptic termini of dopaminergic neurons to facilitate associative learning in *C. elegans*? The dopaminergic system is involved in a wide array of nematode behaviours, including locomotion, egg laying, habituation, defecation and the food-encounter response (basal slowing) (Sawin *et al.*, 2000; McDonald *et al.*, 2006). None of these behaviours is affected by ASIC-1 deficiency (Table I; Figure 4A and B). In addition, dopamine biosynthesis in dopaminergic neurons of *asic-1(ok415)* mutants appears normal (Figure 5). Nonetheless, we found that exogenous dopamine alleviates the learning deficit of these animals (Figure 4C; conditioning to isoamyl alcohol). Conversely, eliminating dopaminergic signalling, either by blocking dopamine biosynthesis or by genetic ablation of dopaminergic neurons (Supplementary Figure 6) recapitulates the associative learning defects of *asic-1(ok415)* mutants (Figure 4C and D; Supplementary Table 1). In both cases, exogenous dopamine supply restores associative learning (Figure 4C and D). Dopaminergic neurons were ablated through ectopic expression of the dominant (*d*), neurotoxic *mec-4(u231)* allele under the control of the *asic-1* promoter

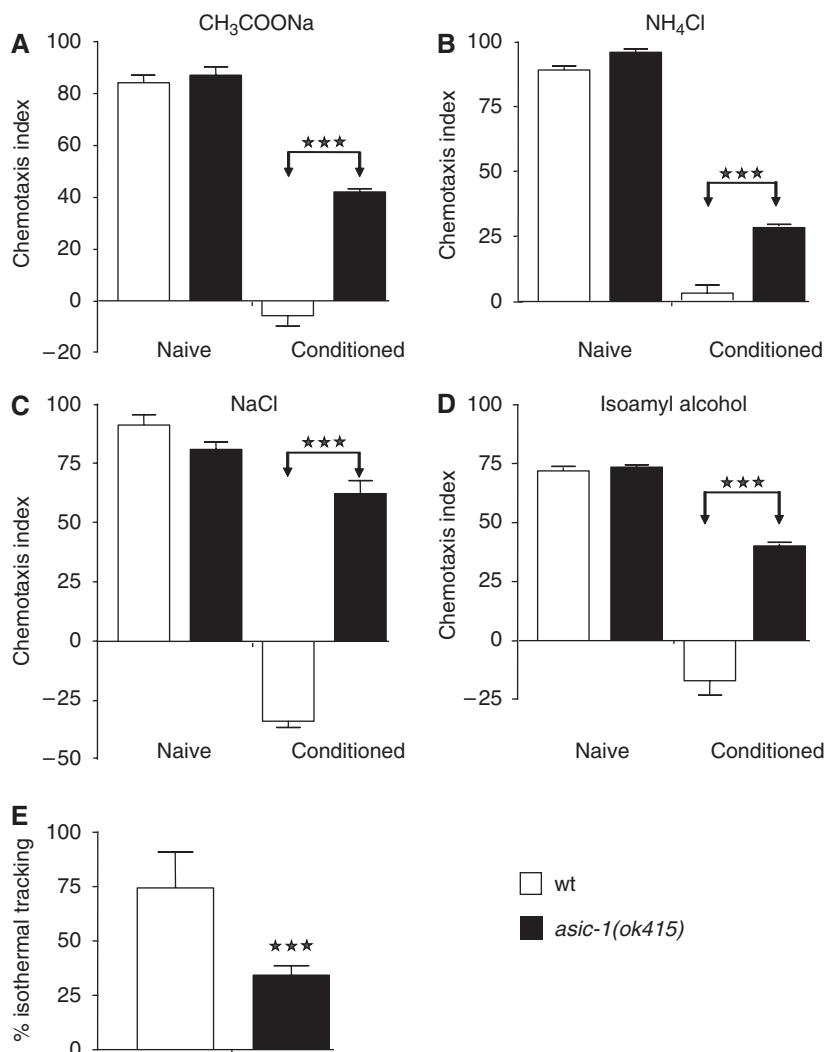


Figure 1 ASIC-1 is required for associative learning in *C. elegans*. (A–D) Conditioning to both gustatory and olfactory cues in the absence of food is impaired in animals carrying a deletion in the *asic-1* gene. Bars depict chemotaxis indices towards the indicated water-soluble (A–C) or volatile compounds (D), calculated for either naive or conditioned wild-type (white) and *asic-1(ok415)* mutant animals (black). Error bars denote standard error of the mean (s.e.m.) values ($n=750$ in four experiments; *** $P<0.001$, unpaired *t*-test). (E) Isothermal tracking performance of *asic-1(ok415)* mutant animals (black bar) is significantly reduced compared with wild type (white bar). Bars depict the percentage of animals that successfully track an isothermal line corresponding to their prior cultivation temperature, in a thermal gradient. Error bars denote s.e.m. values ($n=200$ in eight experiments; *** $P<0.001$, unpaired *t*-test).

(*p_{asic-1mec-4(d)}*), specifically in these cells (Harbinder *et al.*, 1997).

Interestingly, previously characterized dopamine-mediated behaviours such as basal slowing (Sawin *et al.*, 2000) are less sensitive to mild reduction of dopamine, compared with associative learning. By treating *cat-2(e1112)* mutant animals, defective in dopamine biosynthesis, with increasing concentrations of exogenously supplied dopamine, we found that normal conditioning behaviour is restored in *cat-2* mutants at lower concentrations of exogenous dopamine before basal slowing behaviour is back to normal (Supplementary Figure 7). In addition, animals carrying lesions in both D1- and D2-like dopamine receptors (Chase *et al.*, 2004; Suo *et al.*, 2004) show reduced capacity for associative learning (Figure 4E). We obtained similar results with other soluble and volatile chemical cues (not shown). Thus, dopaminergic signalling is required for associative learning in *C. elegans*.

Dopamine release is compromised in *asic-1* mutants

We hypothesized that ASIC-1 functions at dopaminergic synapses to facilitate dopaminergic signalling by augmenting synaptic release of dopamine. We tested this hypothesis in two ways. First, we examined the effect of removing the plasma membrane dopamine transporter (DAT) on associative learning in *asic-1(ok415)* mutant animals. DAT functions to terminate dopamine signalling by mediating dopamine re-uptake at presynaptic termini. After evoked release, extracellular levels of dopamine remain elevated in DAT knockout mice (Gainetdinov and Caron, 2003). In *C. elegans*, the sole plasma membrane DAT is encoded by the *dat-1* gene (Jayanthi *et al.*, 1998; Nass *et al.*, 2005). Deletion of *dat-1* does not alter associative learning in the nematode (Figure 6A). We found that removal of DAT-1 bypasses the requirement for ASIC-1 function in associative learning (Figure 6A). This observation indicates that ASIC-1 contributes to sustain enhanced dopaminergic neurotransmission.

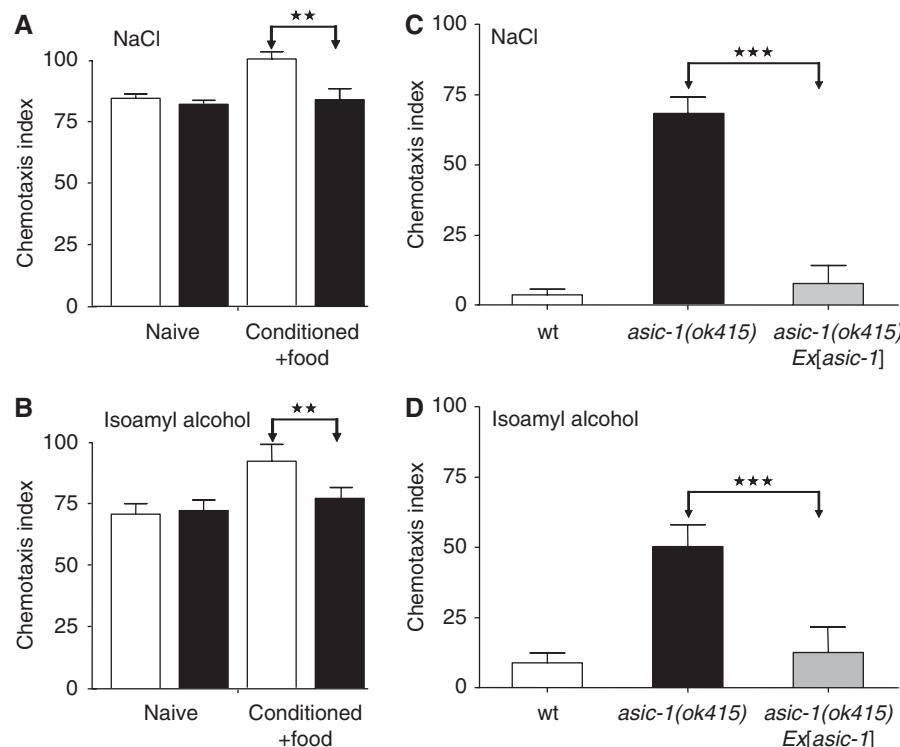


Figure 2 Defective conditioning behaviour of *asic-1(ok415)* mutants is restored by the wild-type *asic-1* gene. (A, B) Chemotaxis to both soluble and volatile chemicals after conditioning in the presence of food. Bars depict chemotaxis indices towards NaCl (A) or isoamyl alcohol (B), calculated for either naive or conditioned wild-type (white) and *asic-1(ok415)* mutant animals (black). Error bars denote s.e.m. values ($n=300$ in two experiments; $**P<0.01$, unpaired *t*-test). (C, D) Complementation of the associative learning deficiency of *asic-1(ok415)* mutant animals by introduction of the wild-type *asic-1* gene. Bars depict chemotaxis indices calculated for animals of the indicated genetic background, after conditioning in the absence of food to NaCl (C) or isoamyl alcohol (D). Error bars denote s.e.m. values ($n=900$ in five experiments; $***P<0.001$, unpaired *t*-test).

Second, we sought to directly compare the activity of dopaminergic synapses in wild-type animals and *asic-1(ok415)* mutants. We monitored synaptic release in dopaminergic neurons of live transgenic animals expressing a synaptobrevin-super ecliptic pHluorin reporter fusion (SNB-1::SEpHluorin) under the *asic-1* promoter, by optical recording of fluorescence recovery after photobleaching (FRAP) (Miesenbock *et al.*, 1998; Samuel *et al.*, 2003). Steady-state levels of SNB-1::SEpHluorin synapse fluorescence are similar between wt animals and *asic-1(ok415)* mutants (Figure 6B). However, we found that the rate of neurotransmitter release at dopaminergic synapses is reduced in *asic-1(ok415)* mutants compared with wild-type controls (Figure 6C). We conclude that ASIC-1 is required to attain normal levels of dopaminergic signalling, necessary for associative learning in *C. elegans*.

ASIC-1 is required for enhanced dopamine release following conditioning

How does training, in the form of conditioning, affect dopaminergic signalling to alter future behaviour? To address this question, we monitored dopamine release before and after conditioning, both in wild-type animals and in *asic-1* mutants. We found that in wild-type animals, conditioning elicits an increase in the rate of dopamine release compared with naive controls (Figure 7A). This observation indicates that dopaminergic neuron synaptic activity is subject to modulation by training in *C. elegans*. We next examined the effect of

conditioning on dopamine release in *asic-1(ok415)* mutant animals. Our analysis revealed that training-induced, enhanced synaptic activity is significantly reduced in *asic-1(ok415)* mutants compared with wild type (Figure 7A). Although, conditioning in these animals does increase the rate of dopamine release above that of naive individuals, synaptic activity remains considerably lower than wild type in both cases. We conclude that ASIC-1 deficiency diminishes the rate of dopamine release at dopaminergic neuron synapses, in both naive and trained animals.

Discussion

ASIC-1 function and dopamine signalling in *C. elegans* associative learning

In this study, we demonstrate that ASIC-1 functions in dopaminergic neurons to mediate normal dopaminergic signalling, necessary for associative learning in *C. elegans*. Our findings reveal a new role of DEG/ENaC ion channels in neuronal communication by regulating the activity of dopaminergic synapses. Biogenic amines such as dopamine are known to modulate behavioural output during operant conditioning in diverse organisms. For example, dopamine is required for aversive reinforcement (punishment response) in *Drosophila* (Riemensperger *et al.*, 2005). In humans, dopaminergic signalling underlies appetitive reinforcement (reward prediction) and addictive behaviour (Pessiglione *et al.*, 2006; Schultz, 2007). Recent studies further underscore the impor-

Table I Phenotypic characterization of *asic-1(ok415)* mutant animals

	Pharyngeal pumping ^a	Defecation ^b	Sinusoidal locomotion wavelength ^c	Sinusoidal locomotion amplitude ^c	Speed ^d	Body size ^e	Gentle body touch response ^f	Harsh touch response ^f	Nose touch response ^f	Egg laying ^g	Thermotactic run ^h
Wild type	223 ± 14	44.1 ± 4.7	0.72 ± 0.1 (0.77 ± 0.3)	0.31 ± 0.2 (0.28 ± 0.3)	1.3 ± 0.6 (2.0 ± 0.8)	1.1 ± 0.4	95 ± 3	98 ± 1	82 ± 2	2.65 ± 0.21	22 ± 2 (25 ± 3)
<i>asic-1(ok415)</i>	218 ± 19	36.6 ± 3.1	0.74 ± 0.2 (0.79 ± 0.4)	0.34 ± 0.2 (0.30 ± 0.4)	1.2 ± 0.7 (2.1 ± 0.9)	1.0 ± 0.2	91 ± 4	96 ± 3	79 ± 3	2.72 ± 0.53	21 ± 3 (26 ± 2)

^aPumps per minute.

^bDuration of defecation cycle in seconds.

^cWavelength and amplitude of sinusoidal track in millimetres. Representative images are shown in Supplementary Figure 5.

^dMillimetres per second.

^eAdult body length in millimetres.

^fPercentage of animals responding.

^gNumber of eggs laid per worm per hour.

^hRun durations in seconds of animals cultivated at 20°C and assayed at 25°C as described earlier (Ryu and Samuel, 2002).

Numbers in parentheses represent measurements after conditioning under starvation. All assays were performed at 20°C unless noted otherwise.

tance of dopaminergic signalling in behavioural plasticity (Day *et al*, 2007; Lammel *et al*, 2008). Similarly, in nematodes, dopamine modulates certain types of behavioural adaptation to distinct sensory cues (Sanyal *et al*, 2004; Kindt *et al*, 2007). Our observations provide insight into a novel mechanism of synaptic plasticity operating at presynaptic terminals of dopaminergic neurons, which links dopaminergic signalling and presynaptic ASICs, to facilitate associative learning in *C. elegans*. Recently, ASICs have been located in mammalian mid-brain dopaminergic neurons (Pidoplichko and Dani, 2006). Further studies should clarify whether these channels mediate associative learning phenomena in mammals, by modulating dopamine signalling in the brain.

ASIC-1 and dopamine modulation of behavioural plasticity

Dopamine signals emitted by the eight dopaminergic neurons of *C. elegans* are received by specific dopamine receptors expressed in many different cell types. Four dopamine receptors (DOP-1 to 4) have been characterized in the nematode (Chase *et al*, 2004; Sanyal *et al*, 2004; Suo *et al*, 2004; Sugiura *et al*, 2005). DOP-1 is a D1-like dopamine receptor expressed in several cells, including mechanosensory neurons, cholinergic motor neurons, interneurons, excretory gland cells, head muscles and neuronal support cells. DOP-2 is orthologous to the human D2-type dopamine receptor and is expressed in all the dopaminergic neurons and some other interneurons. DOP-3 is homologous to mammalian D2 dopamine receptors and is expressed predominantly in neurons other than the dopaminergic neurons and to a lesser extent in body wall muscles. Finally, DOP-4 is similar to D1-like dopamine receptors unique to invertebrates, distinct from mammalian D1-like receptors and is expressed in pharyngeal neurons and interneurons, in the vulva, intestine, rectal glands and rectal epithelial cells.

Which dopamine receptors mediate the effects of dopamine on associative learning? DOP-2 is the dopamine auto-receptor in *C. elegans* and is expressed in dopaminergic neurons. Thus, DOP-2 may modulate dopamine release from these neurons by facilitating neuronal activity, similarly to ASIC-1 (Figure 7B). Consistently with this scenario, we find that DOP-2 deficiency mimics the effects of ASIC-1 deficiency on conditioning phenotypes (Figure 4E). Our results also indicate that DOP-3 functions downstream of DOP-2 because DOP-3 depletion restores normal conditioning behaviour in DOP-2-depleted animals. Interestingly, we also observed that DOP-1 deficiency cancels the effects of DOP-3 deficiency on associative learning (Figure 4E). Indeed, DOP-1 is known to antagonize DOP-3 as *dop-1* mutations suppress *dop-3* mutant phenotypes (Chase *et al*, 2004). DOP-3 and DOP-1 are co-expressed in cholinergic motor neurons and specific mechanosensory neurons (Chase *et al*, 2004). We hypothesize that dopamine facilitates behavioural reprogramming, following training, by modulating the activity of specific neuronal circuits through cognate dopamine receptors. Interestingly, dopamine alters habituation to sudden mechanosensory stimuli by modulating the response of specific mechanoreceptors (Kindt *et al*, 2007). In the absence of food, animals habituate more rapidly than in the presence of food. Such contextual information modulates behaviour in several learning paradigms, in the nematode (Colbert and

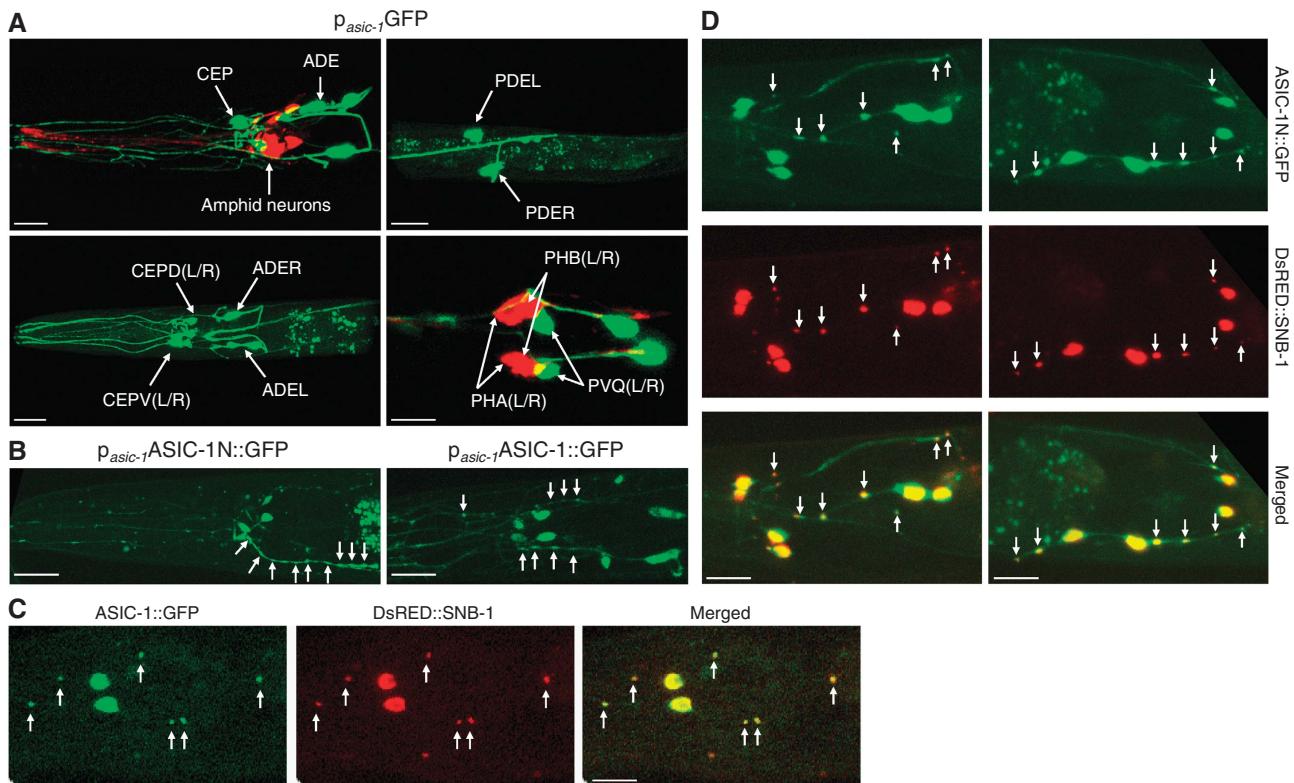


Figure 3 Synaptic localization of ASIC-1 in dopaminergic neurons. **(A)** Images of transgenic animals carrying a *p_{asic-1}*GFP reporter fusion. The *asic-1* gene is expressed in six head neurons shown in the two left panels, the four cephalic (CEP dorsal, ventral/left, right) and two deirid neurons (ADE left, right). The head amphid sensory neurons labelled with the lipophilic carbocyanine tracer Dil (Hedgecock *et al.*, 1985) are shown for reference (top left panel). Two additional deirid dopaminergic neurons in the mid-body section (PDE left, right) express *asic-1* (top right panel). In addition to the eight dopaminergic neurons, *asic-1* is expressed in four tail neurons (PVQ left/right and two more unidentified neurons). The location of phasmid neurons labelled with Dil is shown for reference (bottom right panel). White bars denote 10 µm. **(B)** Images of transgenic animals carrying a full-length *p_{asic-1}*ASIC-1::GFP reporter fusion and an amino-terminal *p_{asic-1}*ASIC-1N::GFP, which includes the complete intracellular amino terminus of ASIC-1, containing the subcellular localization motifs. The punctate distribution of GFP in synaptic areas of head dopaminergic neuron processes is indicated by arrows. White bars denote 10 µm. **(C)** ASIC-1::GFP colocalizes with a DsRED-synaptobrevin fusion, which labels synapses (indicated by arrows), driven by the *asic-1* promoter (*p_{asic-1}*DsRED::SNB-1) in double-transgenic animals carrying both reporter fusions. White bars denote 20 µm. **(D)** Images of double-transgenic animals carrying the amino-terminal *p_{asic-1}*ASIC-1N::GFP reporter together with a *p_{asic-1}*DsRED::SNB-1 fusion. ASIC-1N::GFP (top panels) and DsRED::SNB-1 (middle panels) colocalize in puncta across dopaminergic neuron processes (bottom panel, arrows). White bars denote 20 µm.

Table II Mosaic analysis of *asic-1(ok415)* mutants, expressing the wild-type *asic-1* gene in specific subsets of neurons

Expressing neurons ^a	Chemotaxis index after conditioning ^b
All 12 <i>asic-1</i> -expressing neurons	-2.7 ± 4.8
All 8 dopaminergic neurons	-0.9 ± 5.3
4 non-dopaminergic tail neurons	49.8 ± 4.1
2 ADE dopaminergic neurons	9.8 ± 5.7
4 CEP dopaminergic neurons	20.4 ± 6.2
2 PDE dopaminergic neurons	7.0 ± 4.9

^aMosaic transgenic *asic-1(ok415)*Ex[*p_{asic-1}*GFP]*p_{asic-1}*ASIC-1 animals showing GFP expression in the indicated neurons were grouped and assayed.

^bChemotaxis index (± s.e.m.) towards isoamyl alcohol after conditioning to isoamyl alcohol in the absence of food (*n* = 70 for each mosaic subtype, three independent experiments).

Bargmann, 1997; Rankin, 2000; Law *et al.*, 2004; Kindt *et al.*, 2007). Monitoring the activity of dopamine-targeted neurons in behaving animals should provide insight into how dopamine influences the learning competence of the nervous system in *C. elegans*.

A working model for the involvement of ASIC-1 in modulating dopamine signalling

How does ASIC-1 influence synaptic release in dopaminergic neurons? Our working hypothesis is that ASIC-1 promotes dopamine release at dopaminergic presynaptic termini. According to this hypothesis, ASIC-1 deficiency causes a reduction in the release of dopamine from dopaminergic neurons. We have tested this hypothesis in three ways. First, we examined whether exogenous addition of dopamine reverses the effects of ASIC-1 mutations on conditioning behaviour. Second, we eliminated the plasma membrane DAT-1, which mediates dopamine re-uptake by dopaminergic neurons. DATs function to terminate dopaminergic signalling by clearing dopaminergic synapses and reducing dopamine build-up. Thus, knockout of DAT-1 is anticipated to increase the concentration of dopamine at the synapse and could effectively compensate for ASIC-1 deficiency. Finally, we measured synaptic activity by monitoring the rate of synaptic vesicle exocytosis at dopaminergic synapses using synaptic vesicle-targeted pHluorin and FRET, before and after conditioning, in both wild-type animals and *asic-1(ok415)* mutants. Our findings confirm the original working hypothesis.

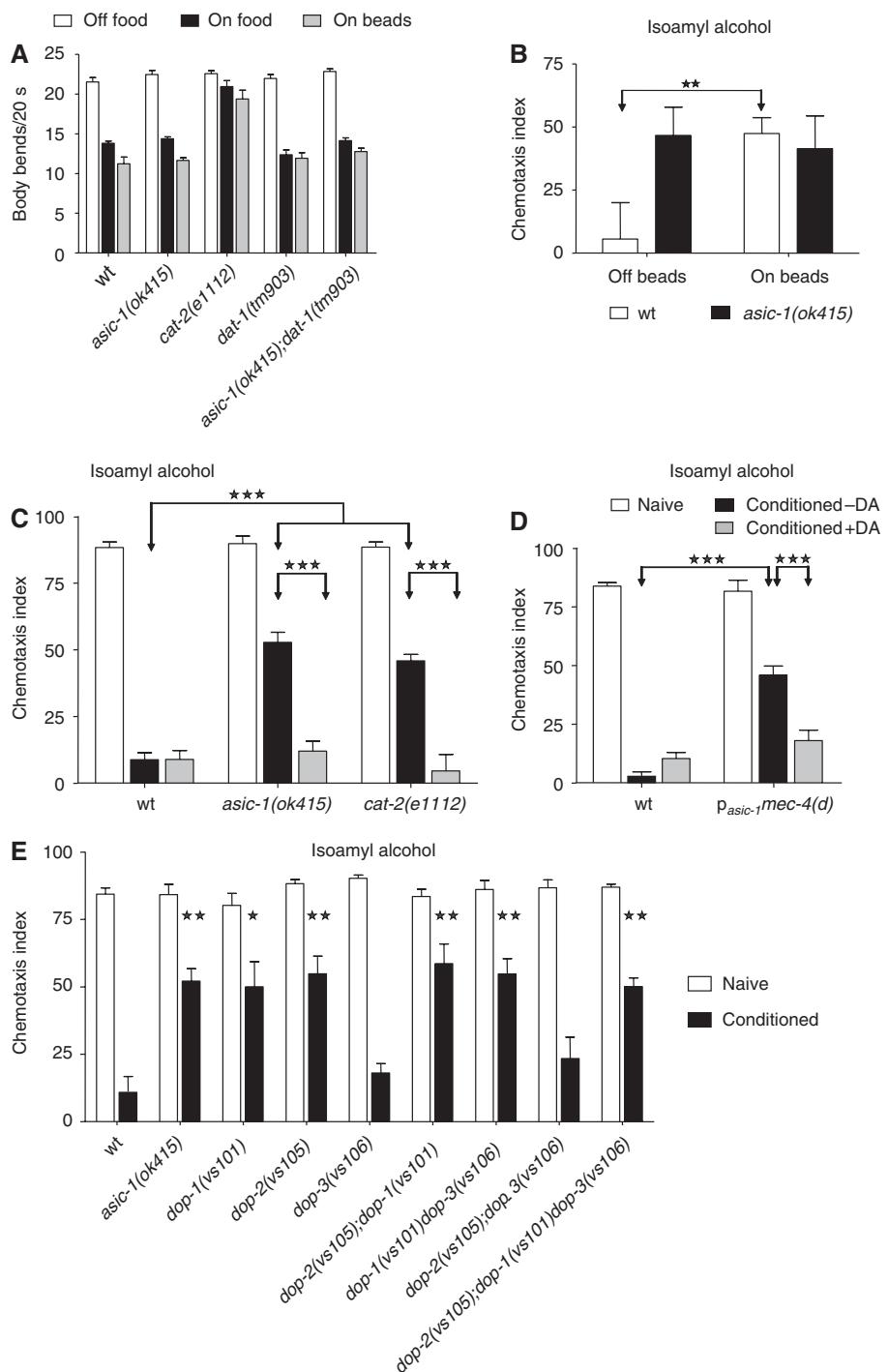


Figure 4 Dopaminergic signalling is required for associative learning in *C. elegans*. (A) *asic-1(ok415)* mutant animals show normal basal slowing behaviour when encountering either food or sepharose beads. *cat-2(e112)* mutants deficient in dopamine biosynthesis do not slow down upon encountering bacterial or sepharose lawns and are included as controls (Sawin *et al.*, 2000). Animals harbouring a lesion in the plasma membrane dopamine transporter gene *dat-1* show normal basal slowing behaviour as do *asic-1(ok415);dat-1(tm903)* double mutants. Error bars denote s.e.m. values ($n=20$ in five experiments). (B) Conditioning in the presence of sepharose beads compromises associative learning in wild-type animals. *asic-1(ok415)* mutants are unaffected. Bars depict chemotaxis indices calculated for animals of the indicated genetic background, after conditioning to isoamyl alcohol. Error bars denote s.e.m. values ($n=500$ in three experiments; ** $P<0.01$, unpaired *t*-test). (C) Exogenous addition of dopamine (DA, 5 mM) restores the capacity for associative learning in *asic-1(ok415)* mutants. Defective dopamine biosynthesis in *cat-2(e112)* mutant animals impairs associative learning, similarly to *asic-1* deficiency. Exogenous addition of dopamine restores associative learning in *cat-2(e112)* mutants. (D) Genetic ablation of dopaminergic neurons results in associative learning defects that are reversed by exogenous addition of dopamine (DA, 5 mM). Animals with at least five ablated dopaminergic neurons, including all CEP neurons were grouped and assayed (see also Supplementary Table 1). For (C, D), bars depict chemotaxis indices towards isoamyl alcohol, calculated for either naive (white bars) or conditioned animals, without (black bars) or with (gray bars) addition of dopamine (DA) in the conditioning medium. Error bars denote s.e.m. values ($n=950$ in five experiments; *** $P<0.001$, unpaired *t*-test). (E) Associative learning in animals lacking specific dopamine receptors. Bars depict chemotaxis indices calculated for animals of the indicated genetic background, before (white) or after (black) conditioning to isoamyl alcohol. Error bars denote s.e.m. values ($n=700$ in four experiments; * $P<0.05$; ** $P<0.01$, unpaired *t*-test).

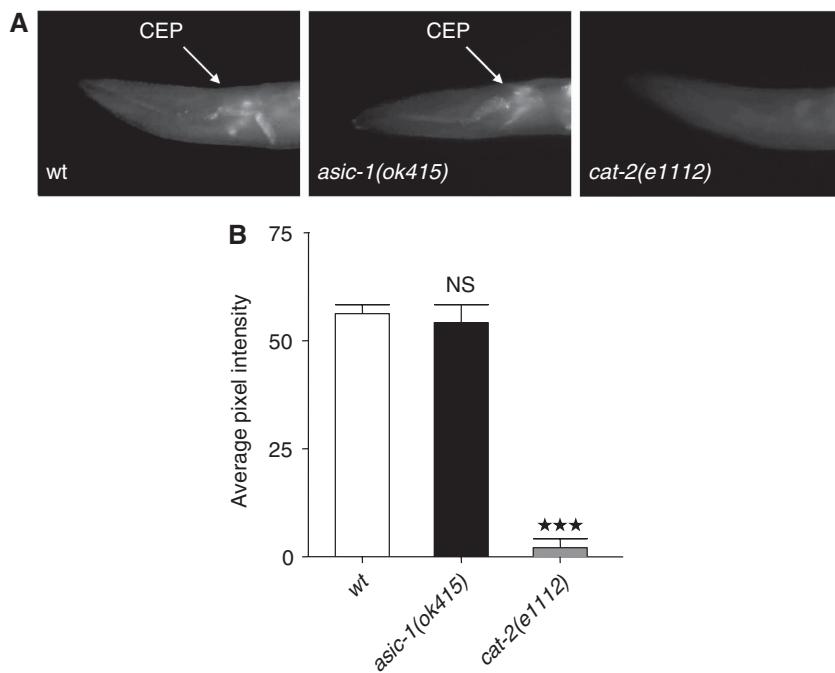


Figure 5 Dopamine biosynthesis is not compromised in *asic-1(ok415)* mutant animals. (A) Dopamine-producing cephalic neurons are visualized by formaldehyde-induced fluorescence in both *wt* and *asic-1(ok415)* mutants. No fluorescence is observed in *cat-2(e1112)* mutants deficient in dopamine biosynthesis, which are included for control. (B) Quantification of formaldehyde-induced fluorescence *wt*, *asic-1(ok415)* and *cat-2(e1112)* animals. Error bars denote s.e.m. values ($n=50$ in three experiments for each genetic background; *** $P<0.001$; NS: difference not significant; unpaired *t*-test, compared with *wt*).

Specifically, we observed that the rate of neurotransmitter release at dopaminergic synapses is lower in animals carrying an *asic-1* deletion. Consistently, DAT-1 deficiency restores normal conditioning behaviour in ASIC-1-deficient animals and exogenous addition of dopamine reverses the effects of *asic-1* deletion. Furthermore, slow dopamine release in *asic-1* mutant animals can be compensated by prolonged exposure to chemoattractants in the absence of food, which reduces their chemotaxis index to wild-type levels (Supplementary Figure 7).

On the basis of the totality of our observations, we propose that ASIC-1 functions to establish a positive feedback loop that reinforces dopamine release upon activation of dopaminergic neurons (Figure 7B). Evoked synaptic vesicle discharge releases both dopamine and protons in the synaptic cleft. Concomitant localized acidification at presynaptic release sites may activate ASIC-1, further depolarizing the presynaptic membrane, thus facilitating dopamine release. This working model bears resemblance to the proposed role of ASICs in LTP through NMDA glutamate receptors at postsynaptic sites in the hippocampus and other brain regions expressing ASICs (Wemmie *et al.*, 2006). Activation of the NMDA receptor or PKC circumvents LTP defects associated with ASIC deficiency, indicating that probably ASIC-1 functions upstream of and normally stimulates NMDA receptor activity (Wemmie *et al.*, 2002). Proton-evoked sodium currents generated by ASIC at postsynaptic terminals are likely to contribute to membrane depolarization and relief of the Mg^{2+} ions blocking NMDA receptors (Wemmie *et al.*, 2002; Gao *et al.*, 2005). Release of protons encapsulated together with neurotransmitters within synaptic vesicles may generate pH fluctuations at synapse micro-domains

during synaptic transmission. Therefore, acidic pH conditions that develop at the synapse as a consequence of synaptic transmission elicit ASIC activation at both pre- and postsynaptic termini, potentiating neuronal communication. Interestingly, ASICs also appear to conduct calcium (Xiong *et al.*, 2004; Yermolaieva *et al.*, 2004). Calcium entry may, in turn, trigger signalling pathways that enhance LTP. Further studies should provide insight into how ASIC-1 activity is regulated to influence dopaminergic signalling during learning.

Materials and methods

Strains and genetics

We followed standard procedures for *C. elegans* strain maintenance, crosses and other genetic manipulations (Brenner, 1974). Nematode rearing temperature was kept at 20°C and the *Escherichia coli* strain OP50 was used as a food source. Some nematode strains were obtained by the *C. elegans* Knockout Consortium (Robert Barstead, Oklahoma Medical Research Foundation, USA), the *Caenorhabditis* Genetics Center (Theresa Stiernagle, University of Minnesota, Minneapolis, USA) and the National Biorepository Project, Japan (Shohei Mitani). The following strains were used in this study: N2: wild-type Bristol isolate; RB680: *asic-1(ok415)*; CB112: *cat-2(e1112)*; *dat-1(tm903)*; *asic-1(ok415)*; *dat-1(tm903)*; LX645: *dop-1(vs100)*; LX702: *dop-2(vs105)*; LX703: *dop-3(vs106)*; LX705: *dop-1(vs100)*; *dop-3(vs106)*; LX706: *dop-2(vs105)*; *dop-1(vs100)*; LX704: *dop-2(vs105)*; *dop-3(vs106)*; LX734: *dop-2(vs105)*; *dop-1(vs100)*; *dop-3(vs106)*; N2Ex[*pasic-1*GFPpRF4], N2Ex[*pasic-1*ASIC-1N::GFPpRF4], N2Ex[*pasic-1*mec-4(u231)p_{myo-2}GFP] referred to in the text as *pasic-1mec-4(d)*, N2Ex[*pasic-1*mec-4(u231)]Ex[*pasic-1*GFP], *asic-1(ok415)*Ex[*pmyo-2*GFP_{asic-1}ASIC-1] referred to in the text as *asic-1(ok415)*Ex[*asic-1*], *asic-1(ok415)*Ex[*pasic-1*GFP_{asic-1}ASIC-1], N2Ex[*pasic-1*ASIC-1N::GFPp_{asic-1}DsRED::SNB-1pRF4], N2Ex[*pasic-1*SNB-1::SEpHluorinpRF4] and *asic-1(ok415)*Ex[*pasic-1*SNB-1::SEpHluorinpRF4].

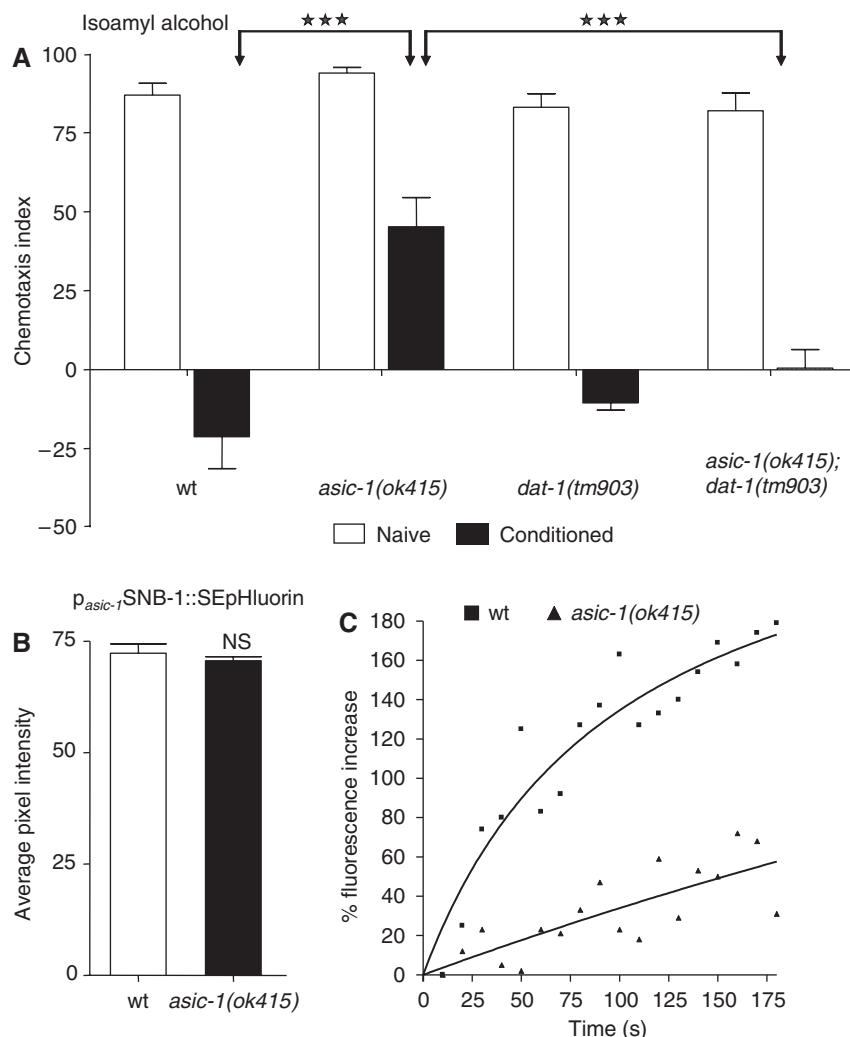


Figure 6 ASIC-1 and dopaminergic signalling. **(A)** Elimination of the plasma membrane DAT-1 dopamine transporter cancels the effects of ASIC-1 deficiency on conditioning behaviour. Associative learning is restored in *asic-1(ok415);dat-1(tm903)* double mutants, carrying lesions in both the *asic-1* and *dat-1* gene. DAT-1 depletion does not affect associative learning. Bars depict chemotaxis indices towards isoamyl alcohol, calculated for either naive (white) or conditioned animals (black) of the indicated genetic background. Error bars denote s.e.m. values ($n = 1100$ in six experiments; *** $P < 0.001$, unpaired *t*-test). **(B)** Steady-state expression of the *pasic-1*SNB-1::SEpHluorin reporter fusion in wild-type and *asic-1(ok415)* mutant animals. Bars depict average pixel intensity calculated from images of SNB-1::SEpHluorin-labelled synapses. Error bars denote s.e.m. values ($n = 150$ synapses in 30 individuals per genetic background; NS: not significant difference compared with wt; unpaired *t*-test). **(C)** Synaptic release at dopaminergic neurons in compromised *asic-1(ok415)* mutants. The rate of fluorescence recovery after photobleaching of synapses labelled with SNB-1::SEpHluorin, specifically in dopaminergic neurons (triangles), compared with wild-type controls (squares). The percentage of fluorescence increase graphed against time. Values are calculated relatively to fluorescence intensity remaining at the point of photobleaching completion.

Behavioural assays

Chemotaxis to soluble and volatile compounds was performed at 20°C, on 9 cm agar plates as described earlier (Ward, 1973; Bargmann and Horvitz, 1991; Bargmann *et al.*, 1993; Colbert and Bargmann, 1995, 1997; Saeki *et al.*, 2001). The chemotaxis index was calculated by subtracting the number of animals found at the trap from the number of animals at the source of the chemical, divided by the total number of animals entered into the assay (Bargmann and Horvitz, 1991). The resulting values were expressed and graphed as percentiles. Both naive and conditioned animals were challenged for 1 h with gradients of NaCl, NH₄Cl, CH₃COONa and isoamyl alcohol (gradient sources: CH₃COONa: 200 mM; NH₄Cl: 250 mM; NaCl: 100 mM; isoamyl alcohol: 1/100 dilution in water). About 200 adult animals for each strain were assayed in each experiment. For conditioning to water-soluble chemicals, animals were exposed to 20 mM NaCl, 50 mM NH₄Cl or 50 mM CH₃COONa on agar plates devoid of bacterial food, for 3 h, prior to assaying chemotaxis to the respective compound. For conditioning to isoamyl alcohol, a 3-μl drop of pure isoamyl alcohol was placed

on the lid of a conditioning plate without food. Animals were conditioned to isoamyl alcohol for 90 min. Conditioning to isoamyl alcohol was also performed on 6 cm assay plates containing 600 μl of 30 mg/ml Sephadex beads. For adaptation assays, animals were exposed to isoamyl alcohol for 180 min in the absence of food. Appropriate time intervals for conditioning and adaptation were determined by time course analysis of chemotaxis after exposure to soluble and volatile cues for periods ranging from 30 to 240 min, in the absence of food. The effects of dopamine on chemotactic behaviours were examined by conditioning both wild-type and mutant animals on agar plates containing 5 mM dopamine hydrochloride (Sigma-Aldrich, St Louis, MO, USA). Dopamine was also added to chemotaxis assay plates at the same concentration. Basal slowing assays with well-fed animals were performed essentially as described earlier (Sawin *et al.*, 2000). For each experiment, 10 young adult animals were followed and their body bends were counted during a 20-s period, both in the absence of food and on a bacterial lawn. Basal slowing was also assayed on thin films of Sephadex G-200 (Sigma-Aldrich) beads on agar plates,

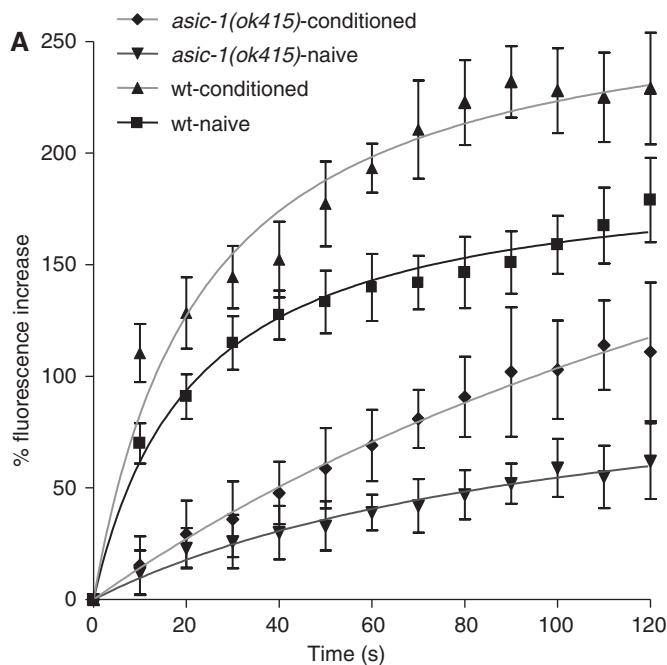
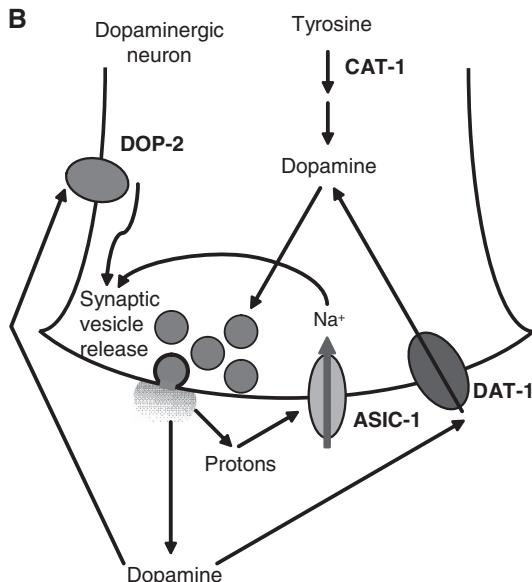


Figure 7 Modulation of dopaminergic signalling by conditioning in wild-type animals and *asic-1(ok415)* mutants. (A) The percentage of fluorescence increase after photobleaching for both naive and conditioned animals of the indicated genetic background is graphed against time. For conditioning, synchronous nematode populations were exposed to isoamyl alcohol in the absence of food. Values are calculated relatively to fluorescence intensity remaining at the point of photobleaching completion. Error bars denote standard deviation ($n > 600$ synapses in 100 individuals). (B) A working model for dopamine signalling modulation at dopaminergic neuron synapses. ASIC-1 functions to facilitate a positive feedback loop that reinforces dopamine release upon activation of dopaminergic neurons. We propose that evoked dopamine release, and the consequent local pH drop, activate the dopamine autoreceptors DOP-2 and ASIC-1, respectively, which promote further dopamine release, resulting in sustained dopaminergic signalling.

used to create axenic substrates with texture simulating that of a bacterial lawn (Sawin *et al.*, 2000). Single worm isothermal tracking assays were performed essentially as described earlier (Hedgecock and Russell, 1975; Mori and Ohshima, 1995; Gomez *et al.*, 2001). In each assay, more than 20 individuals from each genetic background were tested. Thermotaxis index was calculated as the percentage of animals capable of moving for at least half of the assay time on the isothermal line that corresponds to their cultivation temperature, in eight independent assays. Area-restricted search behaviour, osmotic avoidance, nose touch and body touch were assayed as described earlier (Culotti and Russell, 1978; Chalfie and Sulston, 1981; Kaplan and Horvitz, 1993; Hills *et al.*, 2004). We scored sinusoidal locomotion characteristics (amplitude and wavelength) using age-matched animals transferred onto fresh plates and allowed to cut tracks on the bacterial lawn for 10–20 min. The amplitude of the path and the distance between successive peaks in the path (the wavelength) were then measured on photographed tracks. For each strain, 10 trials were conducted and 50 measurements were recorded per trial (Tavernarakis *et al.*, 1997).

Microscopy

Animals were mounted in 2% agarose pads, anaesthetized with 20 mM sodium azide and observed at room temperature. Worms were harvested and observed using a high magnification oil-immersion objective lens (63 \times , Zeiss Plan-NEOFLUAR; numerical aperture 1.4; Carl Zeiss, Jena, Germany), on a confocal microscope (Zeiss AxioScope with a Bio-Rad Radiance 2100 scanhead; Bio-Rad, Hercules, USA), using the Bio-Rad LaserSharp 2000 software package. Labelling of amphid and phasmid neurons with the lipophilic carbocyanine tracer Dil (dioctadecyl-tetramethylindodicarbocyanine-disulphonic acid; Invitrogen, Carlsbad, USA) was done as described earlier (Hedgecock *et al.*, 1985). Labelling of dopaminergic neurons was performed by formaldehyde-induced fluorescence as described earlier (Sulston *et al.*, 1975; Lints and Emmons, 1999). Briefly, young adult animals were treated for 10 min with a solution of 4% paraformaldehyde in phosphate buffer, on a glass slide, at room temperature. After removing excess liquid, animals were incubated for 10 min at 100°C. Fluorescence



was observed using a band-pass 395–440 nm filter for excitation and a 470 nm, long-pass filter for the emission. Measurements of FRAP at *C. elegans* neuronal synapses were done as described earlier (Samuel *et al.*, 2003), using a Zeiss-Bio-Rad Radiance 2100 confocal microscope. Specifically, animals were immobilized on 2% agarose pads with 0.01% levamisole. Individual synapses were visualized by means of Synaptobrevin::SEpHluorin fluorescence. A laser microbeam (wavelength 476 nm, power 5 mW) was used to bleach individual fluorescently labelled synapse regions of all three types of dopaminergic neurons (ADE, CEP and PDE; three consequent photo-bleaching cycles, with duration of 5-s each). Upon completion of photobleaching, fluorescence was reduced by 50–60% of the initial intensity. Subsequently, fluorescence recovery, specifically within the photobleached area, was monitored at a wavelength of 488 nm. At least five synapses for each neuron type were analysed per animal (between 15 and 20 measurements for each individual; more than 10 animals were analysed for each condition/genetic background). Emission intensity was measured on grayscale images with a pixel depth of 8 bit (256 shades of gray). We calculated the mean and maximum pixel intensity for each animal in these images using the ImageJ software (<http://rsb.info.nih.gov/ij/>). For each transgenic line and condition, we processed at least 150 images over at least five independent trials. Colocalization of GFP and DsRED was assessed by merging images obtained sequentially with a 488 nm laser beam and a 515 ± 15 band-pass filter for GFP, and a 543 nm laser beam and a 570 nm long-pass filter for DsRED. Dye filling (Dil; Invitrogen) and visualization of amphid and phasmid nematode neurons was performed essentially as described earlier (Hedgecock *et al.*, 1985; Perkins *et al.*, 1986).

Bioinformatics

All sequences of *C. elegans* genes and proteins used for phylogenetic analysis were retrieved from Wormbase (<http://www.wormbase.org/>; version WS159). The GenBank accession number for the *C. elegans* ASIC-1 amino-acid sequence is AAB93309. BLAST searches (Altschul *et al.*, 1997) were performed with the National Center for Biotechnology Information web servers

(NCBI; <http://www.ncbi.nlm.nih.gov/BLAST/>). Multiple sequence alignments were generated using the M-Coffee algorithm (Notre-dame *et al.*, 2000; <http://tcoffee.vital-it.ch>) and displayed with BoxShade 3.21 (EMBNet; http://www.ch.embnet.org/software/BOX_form.html). Prosites searches for conserved motifs in protein sequences were performed with the Expert Protein Analysis System (ExPASy) proteomics web-based server at the Swiss Institute of Bioinformatics (SIB; <http://www.expasy.ch/tools/scanprosite/>). Transmembrane topology predictions were performed with the Phobius algorithm (Kall *et al.*, 2004); (<http://phobius.cgb.ki.se/>).

Statistics

Chemotaxis and thermotaxis indices, mean and standard error values were calculated from at least five independent experiments. The Student's *t*-test was used for two-way comparisons with a significance cutoff level of $P < 0.05$. Analysis of variance was used for comparisons of multiple groups and was followed by Bonferroni-corrected multiple group comparison *t*-tests. Statistical analysis was carried out using the Microsoft Office 2003 Excel software package (Microsoft Corporation, Redmond, USA) and the Prism software package (GraphPad Software Inc., San Diego, USA).

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Supplementary data

Supplementary data are available at *The EMBO Journal* Online (<http://www.embojournal.org>).

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